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***In utero* exposure to a mixture of the perfluoroalkyl-isopropyl pesticide  
pyrfluquinazon with dibutyl phthalate cumulatively disrupts male rat  
reproductive development via different mechanisms of action**

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## Abstract

Administration of individual chemicals and mixtures during sexual differentiation that disrupt the androgen signaling pathway can induce reproductive abnormalities in male rats. In the current study, we co-administered the heptafluoroisopropyl pesticide pyriproxyfen (PFQ), and dibutyl phthalate (DBP) to pregnant rats during sexual differentiation of the reproductive tract. Both chemicals have been shown to disrupt reproductive tract differentiation in a dose-related manner reducing male anogenital distance (AGD), permanently reducing androgen-dependent tissue weights and sperm counts, and inducing reproductive malformations in male offspring, albeit by different mechanisms of action that converge downstream in the androgen signaling pathway on a common key event. Rats were orally dosed from gestation days 14-18 with dilutions of PFQ and DBP at 0, 12.5, 25, 50, 75 and 100% of the top dose (100 mg/kg PFQ and 750 mg/kg DBP). The mixture ratio was selected such that each chemical would contribute equally to multiple effects on the male offspring reproductive tract and the dose range was designed to determine if the mixture produced additive effects predicted by dose addition or response addition models, or whether significant interactions occurred. Observed data were compared to dose and response addition model predictions. As hypothesized, the mixture reduced F1 male AGD, reproductive organ weights and sperm counts and induced hypospadias with dose addition consistently providing a better prediction of the observed effects than response addition. These results support our hypothesis that chemicals that disrupt the androgen signaling pathway induce dose-additive male reproductive abnormalities regardless of the specific mechanism of action.

Introduction

Per- and polyfluoroalkyl substances (PFAS) vary greatly in structure, and depending on the definition (e.g., OECD, US EPA, etc.), the term “PFAS” may refer to as few as a couple hundred individual chemicals to more than 9,000. The Organization of Economic Cooperation and development (OECD) and the EPA are currently reviewing their definition of PFAS (Wang *et al.* 2021), but currently pyrifluquinazon (PFQ) has been identified as a PFAS by the OECD as well as the USEPA CompTox PFAS Master List ([https://comptox.epa.gov/dashboard/chemical\\_lists/pfasmaster](https://comptox.epa.gov/dashboard/chemical_lists/pfasmaster)) and is one of several registered pesticides with a heptafluoroisopropyl group (Qacemi *et al.* 2019). The structure of this subclass of PFAS is quite different from legacy/straight-chain structures like perfluorooctanoic acid (PFOA) and perfluorooctane sulfonic acid (PFOS), showing the diversity in structure and bioactivity among the PFAS. PFQ is currently registered under the Federal Insecticide, Fungicide and Rodenticide Act (FIFRA) for use on a variety of food crops and ornamental plants and aerial application in the United States (US EPA 2018, 2020).

PFQ and dibutyl phthalate (DBP) have been shown individually to disrupt male reproductive tract differentiation of many of the same tissues in a dose-dependent manner, including reduction of anogenital distance (AGD), androgen-dependent tissue weights and sperm counts (Gray *et al.* 2019a), (Hotchkiss *et al.* 2010; Mylchreest *et al.* 1998; Mylchreest *et al.* 1999; Mylchreest *et al.* 2000). In EPA’s human health assessment of PFQ, the Health Effects Division of the Office of Pesticide Programs selected reduced male rat AGD as the endpoint of concern for females of childbearing age and fetuses. An Acute Population Adjusted Dose (aPAD) of 0.05 mg/kg/day was derived from a developmental NOAEL of 5mg/kg day and a 100-fold UF (10X for inter-species extrapolation, and 10X for intra-species variation).

The current study is similar in design to our fixed ratio binary mixture dilution study with DBP and the androgen receptor (AR) antagonist procymidone (Hotchkiss *et al.* 2010) (Table 1). Herein, we conducted a fixed ratio binary mixture study with PFQ, a perfluoroalkyl-isopropyl pesticide (PFAP), and DBP. To our knowledge, this is the first study showing that a mixture of a PFAS with another chemical produces cumulative, dose additive adverse effects on reproductive development in male rats. There are few, if any, published studies on the effects of PFAS mixed with other chemicals on reproductive development after in utero exposure. A few *in vivo* mixture

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3 studies with PFAS have been published, but these did not compare dose- and response addition  
4 models so it is not possible to determine if the mixture effects were additive, synergistic or  
5 antagonistic (Marques *et al.* 2021; Roth *et al.* 2021).  
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9 One advantage of binary mixture studies, like the current study, is that interpretation of  
10 the nature of the mixture effects is clear. In contrast, it is possible, albeit unlikely, that the effects  
11 in a more complex mixture study could appear dose additive, when in reality, some interactions  
12 in the mixture were antagonistic while others were synergistic. An emerging body of information  
13 supports the hypothesis that when chemicals that disrupt androgen signaling via different  
14 mechanisms of action are administered as a mixture to pregnant female rats during the critical  
15 window of sexual differentiation, permanent adverse effects occur in the offspring at doses lower  
16 than those at which the individual chemicals produce effects, and the mixture effects are best  
17 predicted by dose addition modeling.  
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25 Over the last 20 years, we have conducted several component-based mixture studies with  
26 chemicals that disrupt the androgen signaling pathway in utero via a variety of mechanisms  
27 (reviewed by (Howdeshell *et al.* 2017)). These studies demonstrate that chemicals with diverse  
28 mechanisms of toxicity and modes of action, with different structures, from different chemical  
29 classes produce cumulative effects that are more accurately predicted by dose addition than  
30 response addition. Studies have included binary mixture designs including mixtures of benzyl  
31 butyl phthalate and the pesticide linuron (Hotchkiss *et al.* 2004), DBP and the pesticide  
32 procymidone (Hotchkiss *et al.* 2010), and dipentyl phthalate and the drug simvastatin (Beverly *et al.*  
33 2014) (Table 1), and multichemical mixture studies with 7 (Rider *et al.* 2008), 10 (Rider *et al.*  
34 2010), 15 (Conley *et al.* 2021a) and 18 chemicals (Conley *et al.* 2018). Two of the  
35 multichemical studies (Conley *et al.* 2018; Conley *et al.* 2021b) reported effects at mixture dose  
36 levels well below the individual chemical no-observed effect level on the developing  
37 reproductive system. Taken together, these studies, along with similar studies from other  
38 researchers (Christiansen *et al.* 2008; Hass *et al.* 2007; Kortenkamp 2007; Metzdorff *et al.* 2007),  
39 all indicate that when considering cumulative assessment chemicals should be grouped based  
40 upon their ability to disrupt common toxicological targets as opposed to grouping them by  
41 chemical structure or class, molecular mechanism, or mode of action.  
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In the current study, pregnant rats were exposed by oral gavage to a mixture of PFQ and DBP from gestational days (GD) 14-18, the stage of pregnancy in rat during the critical window of rat sexual differentiation (Wolf *et al.* 2000) (Gray *et al.* 1999) (Carruthers and Foster 2005). The study was a fixed-ratio dilution design and included doses of 0, 12.5, 25, 50, 75 and 100% of the top dose which contained 100 mg/kg for PFQ and 750 mg/kg for DBP. The dose range was designed to determine if the mixture acted in a dose additive, response additive, synergistic (at low doses) or antagonistic (at high doses) manner. Based upon the results of our previous studies, we hypothesized that these two chemicals would act in a dose additive manner because they both disrupt the androgen signaling pathway during sexual differentiation and induce common effects even though they act via different mechanisms of action.

**Methods**

The study consisted of 6 treatment groups administered at 0, 12.5, 25, 50, 75 and 100% of the top dose corresponding to 0, 12.5mg PFQ + 93.75mg DBP, 25mg PFQ + 187.5 mg DBP, 50mg PFQ + 375mg DBP, 75mg PFQ + 562mg DBP, and a top dose of 100mg PFQ + 750mg DBP) with 5 pregnant dams in each group. The top dose of each chemical has been shown to produce a high rate of reproductive tract malformations when administered from GD 14-18 (Gray *et al.* 2019a) (Hotchkiss *et al.* 2010). An examination of the ED<sub>50</sub>s for each chemical indicates that the relative potency factors for PFQ to DBP varies from androgen-dependent endpoint to endpoint. For this reason, PFQ and DBP contribute equally to the development of some abnormalities in the current study, whereas one chemical or the other is primarily responsible for the induction of other effects.

**Animals and administration of chemicals**

Timed-pregnant Sprague Dawley (SD) rats, approximately 90 days old, were purchased from Charles River Laboratories (Raleigh, NC, USA) and shipped to EPA on GD 2 (date of sperm plug positive = GD 1). Animals were housed individually in clear, polycarbonate cages (20 × 25 × 47 cm) lined with laboratory-grade heat treated pine shavings (Northeastern Products, Warrensburg, NY), with a 12:12 light: dark photoperiod (lights off at 18:00) at 20-22 °C and 45-55% humidity. Dams and offspring were provided with NIH07 rat chow during gestation and lactation and offspring were provided NTP 2000 rodent diet after weaning. Rats had access to

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3 filtered (5 micron) municipal tap water *ad libitum* which is tested every 4 months for a subset of  
4 heavy metals, pesticides, and other chemical contaminants and tested monthly for *Pseudomonas*.  
5 These studies were conducted under protocols approved by the National Health and  
6 Environmental Effects Research Laboratory Institutional Animal Care and Use Committee at a  
7 facility accredited by the Association for Assessment and Accreditation of Laboratory Animal  
8 Care (AAALAC).  
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12 Pregnant rat dams were randomly assigned to treatment groups on GD 14 in a manner  
13 that provided each group with similar means and variances in body weight (randomized  
14 complete block design). Dams were weighed and dosed daily by oral gavage at approximately  
15 0730 hr. (EST) from GD 14-18 with the vehicle (laboratory-grade corn oil [CAS #8001-30-7] or  
16 the test substance mixture at 0.25 ml corn oil/kg bodyweight. PFQ was purchased from Chem  
17 Services, Inc. (CAS 337458-27-2, Lot # 2110000, Cat # RPN-13158-5G, purity = 99%) and  
18 DBP was obtained from Research Triangle -Chemical Services for the National Toxicology  
19 Program (Sample Date 12/15/09, RTI Log #: 031609-A-17, Chem ID I16, CAS 84-74-2, Lot #  
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### 30 **Endpoints modeled in the F1 offspring include:**

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33 Anogenital distance (AGD) was measured in 2 day old offspring using a dissecting  
34 microscope with an ocular micrometer with the observer blind to treatment (as previously  
35 described (Ostby and Gray Jr 2004). AGD represents the distance between the base of the  
36 genital papilla and the rostral end of the anal opening. Female-like nipple/areola retention was  
37 evaluated when the pups were 13 days old and the position of the areolae on the milk lines and  
38 the number of areole/nipples were recorded. The age at puberty (as indicated by full preputial  
39 separation- PPS) was examined from 40 days of age, when it was first detected in this study,  
40 until 47 days of age and the age and weight at puberty were recorded. PPS is considered  
41 complete when the glans penis is completely separated from the prepuce (Korenbroet *et al.* 1977).  
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48 Male offspring (total n=120 males) were necropsied as adults and androgen dependent  
49 organ weight changes were measured (glans penis, ventral prostate, seminal vesicles, testes,  
50 epididymis, levator ani/bulbocavernosus muscle, Cowper's glands, kidneys, and liver) and males  
51 were examined for the presence of hypospadias. Other malformations were noted in the higher  
52 dose levels of the mixture, but these were not modeled because we could not determine ED<sub>50</sub> and  
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slope values with any confidence due to the low effect range (0 to 15%) observed. For example, a sensitivity analysis indicated that adding or removing a single affected male from one of the two higher dose groups in the PFQ dose response study changed the actual ED<sub>50</sub> value of 532 mg/kg to 262 mg/kg or 1360 mg/kg, all of which were well above the PFQ content of 100 mg/kg/d in the top dose of the mixture. In addition, these malformations often display very steep slopes with ambiguous logistic regression parameter estimates.

Epididymal sperm counts were also measured. When present, the right epididymis was used to determine caput/corpus and caudal sperm counts as previously described (Gray *et al.* 1989; Gray *et al.* 1995). The right epididymis was cut 13 mm from the tip of the cauda, and the cauda and caput and corpus were placed in separate 20 mL glass scintillation vials containing 2 mL modified M199 media (pre-warmed to 37 °C in an incubator) and minced uniformly with iris scissors. An additional 3 mL of modified M199 were added to each vial after mincing, and the vial was then incubated for approximately 1 hour at 37 °C. At the end of 1 hour, 15 mL of 10% buffered formalin acetate was added to each vial, bringing the total volume to 20 mL (considered to be a 1:20 dilution). The vials are then vortexed for approximately 30 seconds. An aliquot of the diluted sample was then brought to a final dilution of 1:8000 by pipetting 50 µl of the dilution containing minced epididymal tissue and adding 20 mL of 10% buffered formalin acetate to form a second dilution. A homogenous sample of 0.5 mL of the final dilution was counted three independent times using a Beckmann-Coulter cell counter and the average of the three independent counts was used for statistical analyses. The total number of sperm per sample was determined as [(the average of 3 counts) x (16,000)]. This method yields accurate total sperm counts within the normal range of numbers for adult and pubertal male rats (Robb *et al.* 1978) and a low intra-sample coefficient of variation (less than 5%) and low coefficients of variation among control adult male rats (CV control males; PFQ study =19%, DBP study=17%, Mix study = 21%).

## Statistical Methods

Maternal and litter data were analyzed using PROC GLM in SAS 9.4 and each dose group was compared to control using the LSMEANS option. Data collected on multiple F1 offspring per litter were analyzed using litter means values with PROC GLM. Males displaying complete agenesis of an organ were assigned a weight of 0 mg. In addition, males with complete agenesis of the epididymis were assigned a sperm count value of zero.

Dose response data were analyzed using a four-parameter logistic regression (4PL) model in GraphPad Prism 8.4.3 to identify the ED<sub>50</sub> and slope parameters for each variable.

$$R = \frac{1}{1 + \left( \frac{ED50}{D} \right)^p} \quad \text{Equation 1}$$

where  $R$  is the response,  $D$  is the chemical dose,  $p$  is the power or Hill slope of the curve, and  $ED50$  is the exposure dose eliciting a 50 % response. The  $ED50$ s and slopes generated in the individual chemical analyses were used in equations to predict the overall mixture response.

Data were normalized to percent of control in Prism and logistic regression models for most variables were run with the top constrained to 100% and 0% at the bottom, except for AGD at PND2 and testis weights collected at necropsy. The logistic regression model for AGD was constrained to 100% at the top using the mean F1 control male value and the female rat AGD value at the bottom (representing the maximum biologically plausible degree of demasculinization). Since the effect on testis weight is due to the absence or reduction in spermatogenesis due to fluid pressure atrophy that results from epididymal lesions rather than agenesis of the testis, the bottom of the model for paired testes weight was set to the value for paired testis weight typically seen with bilateral testis hypospermatogenesis which was 2.0 g in the PFQ and mixture studies and 1.47 g in the DBP dose response study.

Data from published *in utero* DBP and PFQ studies in our laboratory with GD 14-18 dosing were used to calculate ED<sub>50</sub> and slope parameters for use in dose- (DA) and response- (RA) addition mixture models to predict the effects of the mixture of DBP and PFQ observed in the current study (Hotchkiss *et al.* 2010) (Gray *et al.* 2019a). DA was calculated using two different statistical models. The first model, our laboratory has traditionally used (Rider and



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3 LeBlanc 2005) (Olmstead and LeBlanc 2005) (Rider *et al.* 2008), uses an average slope value to  
4 calculate DA and assumes that the slopes for each chemical in the mixture are similar to one  
5 another for a specific endpoint (referred to herein as DA<sub>1</sub>). The joint toxicity of these binary  
6 mixtures of like-acting chemicals was computed using the following equation (Olmstead and  
7 LeBlanc, 2005):  
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$$R = \frac{1}{1 + \frac{1}{\left( \sum_{i=1}^n \frac{D_i}{ED50_i} \right)^\rho}}$$

Equation 2

where *R* is the response to the mixture, *D<sub>i</sub>* is the concentration of chemical *i* in the mixture, *ED50<sub>i</sub>* is the concentration of chemical *i* that causes a 50% response, and ρ is the power (Hillslope) associated with the chemicals.

Since this assumption is not always met, we also calculated DA using the model described by several authors [for example (Altenburger *et al.* 2000) (Kortenkamp *et al.* 2007) (Metzdorff *et al.* 2007)] and by the NAS (NAS 2008) which does not require parallel slopes for the chemicals in the mixture (referred to as DA<sub>2</sub>). For DA<sub>2</sub>, the dose-additivity predictions were estimated using Equation 3. These are defined by the effect dose of the mixture of DBP plus PFQ for the effect doses of the two chemicals where *p<sub>i</sub>* is the proportion of chemical *i* in the mixture, *EDx<sub>i</sub>* denotes the fractional effect (in %) of the *i*th chemical at *EDx<sub>mixture</sub>*

$$EDx_{mixture} = (p_1/EDx_1 + p_2/EDx_2)^{-1}$$

Equation 3

Response addition (RA) was estimated using Equation 4 where *Ex<sub>mixture</sub>* is the predicted effect of the mixture and *Ex<sub>1</sub>* and *Ex<sub>2</sub>* are the individual effects of DBP and PFQ at a given dose levels.

$$Ex_{mixture} = 1 - [(1 - Ex_1) \times (1 - Ex_2)]$$

Equation 4

The objective of using two DA models was to determine if they provided similar predictions and, if not, to what degree differences in slopes of the dose response curves resulted in differences in DA mixture predictions. We also predicted mixture effects using independent action with a “response addition” (RA) model as described in (Rider *et al.* 2008).

Following these analyses, the “better” mixture model among the two DA models and the RA model was determined by fitting the data with the each 4PL model, constrained to 100% at the top, 0% at the bottom, and to the ED50 and Hillslope value of each model. Following this the Akaike information criterion (AIC) values among the DA<sub>1</sub>, DA<sub>2</sub> and RA models were compared by calculating the probability that one model was more or less informative than another using the equation  $[\exp((AIC_{\min} - AIC_i)/2)]$  which is known as the relative likelihood of model *i* (Burnham and Anderson 2004).

## RESULTS

### Maternal and F1 Effects

Maternal body weight on GD 18 was not significantly reduced at any dose level, being reduced by about 5% in the top dose group, but maternal weight gain was significantly reduced by about 20 g as compared to control dams in the high dose group (Table 2). Postnatal litter sizes were reduced in the top dose group on PND13 and PND23 and the percent of pup mortality was significantly increased in the top dose group by PND23 (Table 2). In addition, pup body weight was significantly reduced on PND 2 in the top dose group by about 1.25 g, approximately 15% of control, but there was no significant effect on male body weight on PND 13 or at weaning.

AGD was not affected in female pups on PND2, whereas AGD in male rat pups was reduced in all dose groups being significant at 25% of the top dose level and above ( $p < 0.0001$ ). The magnitude of the reduction in AGD in males ranged from 10 to 75% of control, being reduced by 45% of control in the 25% of top dose group (control female AGD = 0% of control) (Table 2).

At 13 days of age, control males did not display female-like retained nipples/areolae but treated males displayed nipples/areolae in all dose groups ranging from 0.65 to 12 nipples-areolae/per male, being significant at dose levels of 25% of the top dose and above. The percentage of males displaying any nipples-areolae was significant at all dose levels ranging from 30% affected at 12.5% to 100% in the top two dose groups (Table 2).

The age at puberty could not be evaluated in F1 males in the top two dose groups due to genital malformations and was not significantly affected in males with normal genital morphology (data not shown).

When adult F1 male rat offspring were necropsied, all reproductive organ weights were significantly reduced in the 50, 75% and the top dose group (100%) (Table 3). In the 25% mixture group, males displayed a significant increase in permanently retained female-like nipples, and reduced epididymal sperm. Testicular histopathological results confirmed the observations of gross testicular abnormalities noted at necropsy (Table 4).

Mixture model analyses

We determined the ED<sub>50</sub> and slope values for 12 endpoints for each chemical and used these values to calculate the DA<sub>1</sub>, DA<sub>2</sub> and RA model predictions of joint action of PFQ and DBP binary mixtures (Table 5). These endpoints included AGD, percent of males with any retained female-like nipples/areolae at 13 days of age, number of nipples per males, six androgen-dependent organ weights (ventral prostate, seminal vesicle plus coagulating glands and fluid, levator ani-bulbocavernosus muscles (LABC), testis, Cowper’s glands and epididymal weights), epididymal sperm counts and percent incidence of hypospadias (Figures 1a,b).

When we compared DA and RA models to the observed “best-fit” models using AIC values, we found that DA<sub>2</sub> models produced the most accurate predictions and RA models were the least accurate model for all 11 endpoints (Figure 1 a, b). DA<sub>2</sub> model AIC values also were significantly better than RA predictions for 10 of 11 endpoints (Table 6). DA<sub>2</sub> models were as good as the best fit model for 3 of 11 endpoints. A secondary objective was to compare the fit of two different DA models to the observed mixture data. We found that DA<sub>1</sub> models were equivalent to DA<sub>2</sub> models for 7 of 11 endpoints (Table 6). The ED<sub>50</sub> values with 95% confidence intervals of the observed effects (Table 5) show that the RA models underpredicted the toxicity of the mixture.

Several effects could not be modeled with DA equations due to a lack of significant response from one or both individual chemicals, but the mixture effect exceeded the value predicted by the RA model. For example, the 27% increase in pup mortality versus control (45% versus 18% in the control group) up to weaning in the high dose mixture group exceed the RA

prediction of no increase in pup mortality (0% above control) so it is unclear if this represents an additive or a synergistic outcome.

## DISCUSSION

Results of the current mixture study demonstrate that *in utero* exposure to a PFQ (a perfluoroalkyl pesticide) and DBP (a diortho-phthalate ester) mixture, induces reproductive tract abnormalities in male rat offspring in a dose- rather than response additive manner, even though they act via different molecular mechanisms. These results are consistent with numerous mixture studies our group has conducted over the last 20 years with combined exposure to chemicals that disrupt the androgen signaling pathway during sexual differentiation via different mechanisms of action (reviewed by (Howdeshell *et al.* 2017)) as well as the work of others (Kortenkamp 2020).

The validity of using DA models to predict the *in utero* effects of diverse mixtures of chemicals, including PFQ and DBP, that disrupt androgen signaling and male rat development is supported by an examination of the bioactivity of these chemicals at the pathway level. These chemicals act via adverse outcome pathways (AOPs) with diverse molecular initiating events (MIEs) that converge on a common key event (KE) in an AOP network that regulates development of androgen-dependent tissues (Figure 2). The common KE is dysregulation and reduction of androgen-dependent gene expression in male reproductive tissues. AR antagonists, like vinclozolin or procymidone, accomplish this by blocking androgens from binding to ARs, whereas PFQ has been hypothesized to act as a selective androgen receptor degrader (SARD) (Yasunaga *et al.* 2013), which enhances AR degradation. The phthalates (e.g., di-n-butyl phthalate, di(2-ethylhexyl) phthalate, dipentyl phthalate, butyl benzyl phthalate, di-isobutyl phthalate) reduce fetal testis testosterone production reducing the levels of androgens available to the cell/receptor. Chemicals like finasteride inhibit type II and III 5 $\alpha$  reductase, the enzyme in tissues that converts testosterone to dihydrotestosterone (a more active androgen that has higher affinity for the AR). All these molecular perturbations reduce AR/AR homodimer levels, androgen response element (ARE) DNA activation, androgen-dependent mRNA and protein synthesis levels and growth and differentiation of androgen-dependent tissues in the fetus. As a result, male offspring display agenesis or hypoplasia or malformations of androgen-dependent tissues. In addition, we have found that a mixture of DBP and 2,3,7,8-TCDD (dioxin), an AhR

agonist, disrupted differentiation of several reproductive tissues in a manner that exceeded response addition (Rider *et al.* 2010). In summary, an examination of the events disrupted in the androgen signaling pathway by DBP and PFQ at the cellular-molecular level explains why they behave in a dose additive manner. Overall, it is not important to an androgen-dependent cell what MIE reduced the androgen signal to the receptor, it is only important that the signal is reduced.

As stated above, PFQ has been hypothesized to disrupt AR function by enhancing receptor degradation (Yasunaga *et al.* 2013) as opposed to acting as a competitive inhibitor of androgen binding to AR. Although the focus on the development of drugs to treat prostate cancer has been on competitive antagonists like flutamide that compete with androgens for binding to the AR ligand binding domain (LBD), many efforts have shifted to developing drugs that target the N-terminal domain (NTD) rather than the LBD. These include chemicals that function as SARDs or inhibit nuclear AR localization by interacting with specific sites on the NTD (Mohler *et al.* 2021; Narayanan *et al.* 2008). While PFQ may function as a SARD, PFQ also does antagonize the effects of dihydrotestosterone *in vitro*, albeit at high concentrations (Gray *et al.* 2019a). Furthermore, PFQ is extensively metabolized, so it is possible that one of the more than a dozen metabolites acts as a competitive AR antagonist rather than PFQ acting as a SARD.

In contrast to PFQ, DBP and MBP, the active metabolite of DBP, do not interact with the AR *in vitro* (Parks *et al.* 2000) or *in vivo* (Lee and Koo 2007). DBP is one of several diortho-phthalate esters that act as reproductive toxicants *in utero* in the rat by disrupting fetal testis gene expression (Gray *et al.* 2021; Hannas *et al.* 2011) and hormone production (Furr *et al.* 2014) sufficiently to induce permanent reproductive tract abnormalities in F1 male offspring known as the “Phthalate Syndrome” (Foster 2006). Since most people including pregnant women, are exposed to one or more phthalates like DBP (Adibi *et al.* 2008) (Zota *et al.* 2014), any human exposure to PFQ would also very likely be concurrent with exposure to phthalates and other ubiquitous contaminants.

PFQ is classified by several agencies, including the OECD and USEPA, as a PFAS because PFQ is one of a number of perfluoroalkyl pesticides that contain a heptafluoroisopropyl group. However, PFQ has a notably different structure from legacy/straight-chain PFAS like PFOS and PFOA or perfluoroalkyl ether PFAS like HFPO-DA (GenX) (Figure 3). Similar to other pesticides, and unlike many PFAS, there is an extensive data base on PFQ toxicity, fate, and

transport. In the last few years, the numbers of PFQ-like hepta- and hexa-fluoroisopropyl PFAPs registered or under development have been increasing because addition of this functional group has been reported to provide many traits considered to be beneficial for pesticides, including increased uptake, enhanced insecticidal toxicity and persistence (Qacemi *et al.* 2019). Examples include other heptafluoroisopropyl PFAPs include brofranalide [(registered by EPA early in 2021) (<https://www.regulations.gov/docket/EPA-HQ-OPP-2018-0053/document>)], and pyflubumide [(residue levels established by EPA in late 2021) (<https://www.regulations.gov/search?filter=pyflubumide%20>)]. In 2016 EPA stated that it intended to cancel the registration of four pesticides containing flubendiamide, concluding that the continued use would result in unreasonable adverse effects on the environment, particularly benthic invertebrates (<https://www.epa.gov/ingredients-used-pesticide-products/flubendiamide-notice-intent-cancel-and-other-supporting>, accessed 12/30/2021). In addition, the fungicide ZJ5337 (used to control rice blast in China (Hu *et al.* 2014) also contains a heptafluoroisopropyl group. Taken together, the literature indicates that there are a number of PFAP pesticides in use or under development indicating that PFQ is not unique in this regard. However, for those PFAP pesticides with available summary assessments (regulations.gov), the data suggest that the presence of a hepta- or hexa-fluoroisopropyl side chain does not necessarily confer PFQ-like antiandrogenic bioactivity *in vivo*.

In addition to the above PFAPs, there are PFAPs and inert ingredients in pesticides with structures that are similar to PFOA and PFOS that have been used in the US and in other countries including sulfuramid, flusulamid, LPOS, and SIOC-I-013. Examination of the risk assessment summaries provided in EPA documents (in Regulations.gov) for the four aforementioned PFAPs does not indicate that any of these have reproductivity toxicity profiles like PFQ.

Although PFQ is the only PFAS that we are aware of that target development of the male reproductive tract via AR disruption *in vivo*, a number of *in silico* and *in vitro* studies provide a different perspective. These studies have reported that a considerable number of PFAS interact with the ligand binding domain of AR (Di Nisio *et al.* 2019; Yu *et al.* 2022). Interestingly, a number of these PFAS also contain perfluoroisopropyl side chains, like PFQ (Azhagiya Singam *et al.* 2020; Tachachartvanich *et al.* 2022). Furthermore, Kjeldsen et al. (Kjeldsen and Bonefeld-Jorgensen 2013) reported that a mixture of five PFAS displayed antiandrogenic activity *in vitro*

that was additive at the 95% and 90 % levels but appeared synergistic at the 85% of control level. Because *in silico* and *in vitro* assays of AR bioactivity have a high false positive rates (Gray *et al.* 2020) (Charles *et al.* 2005; Fabian *et al.* 2018; Sonneveld *et al.* 2006) these observations would need to be confirmed *in vivo* in (anti)androgen screening assays (EPA 2009; Gray *et al.* 2005) and/or *in utero* (Ostby and Gray Jr 2004) to determine if any of these PFAS have the potential to cause adverse reproductive effects via disruption of the androgen signaling pathway.

As discussed above, the validity of using DA models rather than RA models to predict the *in utero* effects of diverse mixtures of chemicals that disrupt androgen signaling and male rat development is supported by an examination of the bioactivity of these chemicals at the cellular-molecular level. All these chemicals act via adverse outcome pathways (AOPs) that converge on a common key event (KE) in an AOP network that regulates the sequence of molecular events in cells that participate in the development of the androgen-dependent tissues.

In summary, a DBP and PFQ mixture behaved in a dose additive manner because these two chemicals target common reproductive tissues via disruption of overlapping AOPs that converge on a common KE in a fetal androgen signaling AOP network (Figure 2). They act via different MIEs and MOAs but they both reduce the hormone signal at the AR, thus reducing androgen-dependent mRNA, and protein synthesis during fetal life, which in turn induces permanent reproductive abnormalities. Overall, an examination of our mixture studies and similar studies in the literature on the effects of antiandrogens on male reproductive tract development consistently indicate: 1) Dose addition should be considered as the default model for assessing the effects of mixture composed of chemicals with different mechanisms that disrupt common effects regardless of whether or not they share a common MIE, 2) RA models can grossly underestimate the hazard of a mixtures of chemicals, and 3) Failure to account for the effects of mixtures during development and other life stages is not fully protective and could potentially lead to adverse consequences for human and ecosystem health.

### Figure Legends.

Figure 1 a,b. A comparison of the two dose addition ( $DA_1$  and  $DA_2$ ) and the response addition (RA) models with the observed effects of in utero administration of the mixture of dibutyl phthalate (DBP) and the PFAS pesticide pyriproxyfen (PPF) on androgen-dependent tissues in the male rat offspring. The major difference between  $DA_1$  and  $DA_2$  is that  $DA_1$  assumes that the two chemicals have similar dose response slope parameters whereas  $DA_2$  does not.  $DA_2$  was generally the better model among the three models.

Figure 2. An adverse outcome pathway (AOP) network showing the two AOPs disrupted via different molecular initiating events and different AOPs that converge on a common key event in the AOP network.

Figure 3. Two dimensional structures of dibutyl phthalate (DBP) and pyriproxyfen (PPF) (on the left) compared to linear PFAS found in some drinking and surface waters.



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Table 1  
Binary Mixture Studies Summary  
Administered by Oral Gavage Gestational Days 14 to 18

Chemical A			Chemical B			A+B=?			Effect
Name	Dose mg/kg/d	MOA/MIE	Name	Dose mg/kg/d	MOA/MIE	Chemical A % Malformed	Chemical B % Malformed	Chemical A+B % Malformed	
Vinclozolin	50	AR antagonist	Procymidone	50	AR antagonist	0	0	54	Vaginal Pouch Hypospadias
						10.2	0	96	
DBP	500	Fetal Testis Insl3 and Androgen Levels	Procymidone	50	AR antagonist	0	0	26.7	Vaginal Pouch Hypospadias Gubernaculum Testis Agenesis
						0	1.5	48.8	
						0	0	29	
DBP	500	Fetal Testis Insl3 and Androgen Levels	BBP	500	Fetal Testis Insl3 and Androgen Levels	0	0	40	Vaginal Pouch Hypospadias Gubernaculum Testis Agenesis Epididymal Agenesis
						0	0	50	
						0	8	100	
						9	23	100	
DBP	500	Fetal Testis Insl3 and Androgen Levels	DEHP	500	Fetal Testis Insl3 and Androgen Levels	3	0	25	Hypospadias Gubernaculum Testis Agenesis Epididymal Agenesis
						3	27	75	
						26	5	65	
BBP	500	Fetal Testis Function	Linuron	75	Fetal Testis Androgen Levels AR antagonist	0	0	41	Vaginal Pouch Hypospadias Epididymal Agenesis Seminal Vesicle Agenesis
						0	0	56	
						12	62	97	
						12	13	62	
DBP	500	Fetal Testis Insl3 and Androgen Levels	2, 3, 7, 8 TCDD	0.0013 to 0.002	AhR Agonist	24	0	45	Epididymal Agenesis or Atrophy Epididymal Sperm % Reduced Epididymal aspermia Seminal Vesicle Vas Deferns Agenesis
						7	21	37	
						9	0	21	
						5	0	28	
						14	0	26	
DBP	Fixed ratio dilution study 94 to 750	Fetal Testis Insl3 and Androgen Levels	Pyrifluquinazon	Fixed ratio diultion study 12.5 to 100	AR degradation	See Current Study Results			

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**Table 2. Litter and neonatal effects of *in utero* administration of a Dibutyl Phthalate and the PFAS pesticide Pyrifluquinazon**

Percent of Top Dose	0	12.5	25	50	75	100
Pyrifluquinazon Dose (mg/kg/d)	0	12.5	25	50	75	100
Dibutyl Phthalate Dose (mg/kg/d)	0	93.75	187.5	375	562	750
Number of Dams	5	5	5	5	5	5
Implantations	13.6 ±1.3	12.8 ±0.2	13.4 ±0.6	14.2 ±0.6	13.6 ±0.2	12.4 ±1.2
GD 18 Maternal Body Weight (g)	354.7 ±15.5	354.2 ±4.9	358.3 ±6.4	361.8 ±4.0	364.2 ±8.7	337.7 ±10.1
GD 14 to 18 Maternal Weight Gain (g)	42.2 ±3.4	<b>30.2 ±1.1 *</b>	33.8 ±0.7	35.0 ±1.9	<b>30.5 ±3.2 *</b>	<b>20.1 ±7.1 **</b>
Litter Size PND 2	13.4 ±1.3	12.0 ± 0.5	13.0 ±0.7	13.4 ±0.9	11.2 ±1.0	10.8 ±1.3
Female Pup weight (g) PND 2	7.82 ±0.10 (35)	7.93 ±0.27 (29)	7.45 ±0.12 (39)	7.51 ±0.09 (39)	7.52 ±0.14 (26)	<b>6.72 ±0.13(29) **</b>
Male Pup weight (g) PND 2	8.20 ±0.12 (32)	8.23 ±0.14 (30)	7.94 ±0.09 (36)	7.79 ±0.11 (28)	7.60 ±.12 (30)	<b>6.93 ±0.13 (22) **</b>
Female AGD (mm)	1.54 ±0.03 (35)	1.65 ±0.04 (29)	1.51 ±0.3 (30)	1.42 ±0.03	1.47 ±.03 (26)	1.59 ±0.03 (29)
Male AGD (mm)	3.71 ±0.06 (32)	3.46 ±0.07 (30)	<b>2.78 ±0.04 (36) **</b>	<b>2.51 ±0.08 (28) **</b>	<b>2.15 ±0.04(30) **</b>	<b>2.18 ±0.07 (22) **</b>
Male AGD (% of Androgen Effect)	100 ±2.82	88.1 ±3.3	55.6 ±1.98	42.9 ±3.6	26.1 ±1.9	27.4 ±3.2
% Pup Loss (Implants - PND23 litter size)	18.3 ±8.2	15.6 ±2.4	15.8 ±10.4	19.2 ±3.6	26.7 ±5.7	<b>45.2 ±8.9 *</b>
Retained Nipples-Areolae/Male PND 13	0 ± (29)	0.6 ± (29)	<b>4.7 ±0.4 (34) **</b>	<b>6.6 ±0.4 (22) **</b>	<b>11.7 ±0.2 (27) **</b>	<b>12 ±0 (17) **</b>
% Males with any Retained Nipples-Areolae PND 13	0 ±0	<b>30.0 ±0.09 **</b>	<b>91.2 ±0.05 **</b>	<b>100 ±0 **</b>	<b>100 ±0 **</b>	<b>100 ±0 **</b>

Footnotes. F1 data were analyzed using litter means including body weight, AGD, nipples-aerolae, survival and preputial separation

F1 means ± standard error (sample size) are based upon individual pup values, not litter means

\*\* p <0.01

\*p <0.05

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**Table 3. Adult F1 male rat organ weights, gubernaculum testis length (paired), number of retained female-like nipples/areolae and epididymal sperm counts.**

Percent of Top Dose	0			12.5			25			50			75			100		
F1 Male Offspring	Mean	SE	N	Mean	SE	N	Mean	SE	N	Mean	SE	N	Mean	SE	N	Mean	SE	N
Body Weight at Necropsy (g)	745.5	17.79	21	718.3	15.01	21	702.5	15.49	21	685.6	16.54	20	696.3	32.99	15	<b>667.7**</b>	22.97	20
Glans Penis (mg)	141.9	1.82	21	134.8	1.94	21	<b>128.9 *</b>	2.80	19	<b>126.7**</b>	4.04	12	<b>97.8*</b>		1	<b>82.9**</b>		1
Levator Ani-bulbocavernosus (g)	1.76	0.05	21	1.61	0.03	21	1.51	0.05	21	<b>1.06**</b>	0.05	20	<b>0.71**</b>	0.04	15	<b>0.49**</b>	0.06	20
Ventral Prostate (mg)	673.8	42.47	21	556.1	30.10	21	531.8	34.11	21	<b>386.0**</b>	48.52	20	<b>56.4**</b>	40.61	15	<b>0**</b>	0.00	20
Seminal Vesicle (g)	2.07	0.06	21	1.91	0.05	21	2.01	0.05	21	<b>1.73*</b>	0.11	18	<b>0.16**</b>	0.09	15	<b>0.02**</b>	0.02	20
Paired Testes (g)	4.24	0.07	21	4.47	0.06	21	3.86	0.08	21	<b>3.77**</b>	0.11	20	<b>2.20**</b>	0.19	15	<b>2.46**</b>	0.22	20
Gubernaculum Testis (mm)	19.2	0.46	21	18.1	0.61	21	15.6	0.48	21	19.4	0.97	20	<b>110.0**</b>	23.30	15	<b>161.8**</b>	14.26	20
Paired Epididymides (mg)	1460.7	26.24	21	1495.1	20.26	21	1305.3	39.36	21	<b>1287.3*</b>	53.24	20	<b>121.8**</b>	51.28	15	<b>1.3**</b>	1.29	19
										*								
Cauda Epididymis (mg)	309.9	7.10	21	307.2	9.10	21	<b>281.4*</b>	7.50	21	<b>274.9**</b>	11.30	20	<b>23.5**</b>	20.50	15	<b>1.23**</b>	0.00	19
Epididymal sperm count x10 <sup>6</sup>	177.1	9.65	15	179.3	6.92	16	<b>135.2*</b>	13	16	<b>142.6*</b>	14.5	16	<b>7.83**</b>	2.3	3	<b>1.55**</b>		1

Footnotes

N is the number of individual males, analyses for dose effects used Litter Means, organ weights were not taken on all males but they were all examined for testis and epididymal histopathology

Organ weights were not taken on all males but they were all examined for testis and epididymal histopathology

**\*\* p<0.01**  
**\*p<0.05**

Table 4. Dose related Reproductive alterations in F1 Male Rat Offspring after in utero exposure to a mixture of dibutly phthalate and pyrifluquinazon

Percent of Top Dose of the Mixture Abnormalities	0 Mean	SE	N males	12.5 Mean	SE	N males	25 Mean	SE	N males	50 Mean	SE	N males	75 Mean	SE	N males	100 Mean	SE	N males
Agenesis of the Coagulating Glands	0	0	28	0	0	27	0	0	31	0	0	19	52.4**	11.2	21	50**	11.5	20
Epididymal Agenesis*	0	0	28	0	0	27	3.2	3.2	31	15.8*	8.6	19	100**	0	21	85**	8.2	20
Epididymal Histological Hypospermia	0		28	0		27	3.2		31	5.3			75.0		3	no sample		0
Gubernaculum Testis Agenesis	0	0	28	0	0	27	0.0	0.0	31	0	0	19	47.6**	11.2	21	90**	6.9	20
Hypospadias	0	0	28	0	0	27	6.5	4.5	31	47.4**	11.8	19	100**	0	21	100**	0	20
Any Major Abnormality	3.6	3.6	28	3.70	3.70	27	16.1	6.7	31	57.9**	11.6	19	100**	0	21	100**	0	20
% of Males with and Nipples	0	0	28	3.70	3.70	27	64.5**	8.7	31	68.4**	11.0	19	100**	0	21	100**	0	20
Seminal Vesicle Agenesis	0	0	28	0	0	27	0	0	31	10.5	7.2	19	61.9**	10.9	21	90**	6.9	20
Testis Abnormalities	3.6	3.6	28	0	0	27	9.7	5.4	31	15.8	8.6	19	100**	0	21	100**	0	20
Undescended Testis	0	0	28	0	0	27	0	0	31	0	0	19	23.8**	9.5	21	75**	9.9	20
Unilateral or Bilateral Testis Tubular Atrophy	3.6		28	0		27	9.7		31	11.3		19	100		21	100		20
Vaginal Pouch	0	0	28	0	0	27	0	0	31	10.5	7.2	19	90.5**	6.6	21	90**	6.9	20
Vas Deferens Agenesis	0	0	28	0	0	27	0	0	31	5.3	5.3	19	52.4**	11.2	21	70**	10.5	20
Ventral Prostate Agenesis	0	0	28	0	0	27	0	0	31	5.3	5.3	19	66.7**	10.5	21	90**	6.9	20

N is the number of individual males, analyses for dose effects used Litter Means

\*\* p<0.01

\*p<0.05

\*Agenesis refers to organs that display gross partial or complete agenesis or hypoplasia at necropsy

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**Table 5. Results of four parameter logistic (4PL) regression modeling of the individual dibutyl phthalate (DBP) and pyrifluquinazon (PFQ) (two columns on the right) and the two dose addition (DA) and response addition models fit to the observed mixture data. DA1 and DA2 4PL models were calculated using Equations 2 and 3, respectively, in the described in the methods.**

		Observed	DA2	DA1	RA	DBP	PFQ
Anogenital distance	ED50	30.36	43.68	47.31	48.18	829.6	70.95
	CI	24.48 to 37.15					
	HillSlope	1.634	1.497	1.859	1.67	3.408	1.451
Ventral Prostate Weight	ED50	40.92	42.88	44.78	67	987	67.15
	CI						
	HillSlope	2.697	3.361	2.478	2.752	13.5	2.734
Seminal Vesicle Weight	ED50	56.76	52.1	48.29	95.99	737.9	111.4
	CI	50.87 to 63.32					
	HillSlope	8.948	9.746	52.32	12.02	39.55	5.247
Testis Weight	ED50	54.98	53.59	50.96	98.32	755.0	115.0
	CI	50.87 to 63.32					
	HillSlope	3.758	10.07	60.89	8.099	26.30	5.89
Testis Tubular Semiferous Hypospermatogenesis	ED50	52.28	46.56	46.54	68.89	550	127.70
	CI	ambiguous					
	HillSlope	29.96	3.407	3.339	3.945	3.791	2.89
Epididymal Weight	ED50	56.93	51.43	46.53	76.91	579.4	153.9
	CI	52.41 to 61.83					
	HillSlope	8.513	6.615	8.011	6.815	6.725	6.431
LABC Weight	ED50	60.17	62.6	62.92	85.79	967	122.8
	CI	51.10 to 70.81					
	HillSlope	1.878	2.136	2.363	1.993	2.966	1.707
% OF 12 Nipples/ Areolae per male	ED50	34.45	30.52	31.34	41.33	954	41.33
	CI	28.34 to 41.48					
	HillSlope	2.36	2.308	2.295	1.734	25	1.734
% males with any Nipples/Areolae	ED50	15.09	15.08	14.41	18.52	584.7	19.05
	CI	13.41 to 16.99					
	HillSlope	4.498	4.839	3.142	8.114	1.638	9.203
HYPOSPADIAS	ED50	50.3	38.27	38.73	54.54	1000	54.54
	CI	47.81 to 52.80					

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	HillSlope	17.64	3.668	34.07	2.607	64	2.607
Epididymal Sperm Count	ED50	57.07	52.63	52.64	75.74	587	160.9
	CI	46.51 to 70.02	1.721	1.721	4.41		
	HillSlope	10.71	4.067	3.96	4.4	4.27	3.710
Cowper's Gland Weight	ED50	67.84	54.07	56.26	90.71	1084	93.81
	CI	48.50 to 94.89					
	HillSlope	15.44	1.299	3.146	0.929	8.2	0.909

CI - 95% confidence interval

DA1 - Dose Addition Prediction model 1

DBP - Dibutyl Phthalate

DA2 - Dose Addition Prediction model 2

PFQ - Pyriproxyfen

RA - Response Addition

ALL 4 PL MODELS

Table 6. A summary of the preferred models is shown in the lower right panel (based upon p<0.05 of the AIC values)

AIC Table		
Anogenital Distance		
AIC DA OBS	143.4	a
AIC DA2	162.8	b
AIC_RA	170	c
AIC_DA1	170.7	c
% Of Males With Any Nipples		
AIC DA OBS	998.6	a
AIC DA2	1005	b
AIC_DA1	1005	b
AIC_RA	1031	c
% OF 12 NipplesS		
AIC DA OBS	142.6	a
AIC_DA1	144.6	a
AIC DA2	145.7	a
AIC_RA	157.2	b
Testis Weight		
AIC DA OBS	159.6	a
AIC DA2	188.3	b
AIC_DA1	188.4	b
AIC_RA	202.1	c
Ventral Prostate Weight		
AIC DA OBS	179.5	a
AIC DA2	179.7	a
AIC_DA1	180.8	a
AIC_RA	196.5	b
LABC Weight		
AIC DA OBS	139.3	a
AIC DA2	140.6	a
AIC_DA1	141.7	a
AIC_RA	151.2	b
Epididymal Weight		
AIC DA OBS	141.5	a
AIC DA2	156.8	b
AIC_DA1	179.3	c
AIC_RA	190.5	d
Seminal Vesicle Weight		
AIC DA OBS	157.3	a
AIC DA2	164.1	b
AIC_DA1	203.3	c
AIC_RA	223.4	d
Cowper's Gland Weight		
AIC DA OBS	176.3	a
AIC_DA1	199.7	b
AIC DA2	212.4	b
AIC_RA	215.6	b
Epididymal Sperm Count		
AIC DA OBS	442.6	a
AIC DA2	452.6	b
AIC_DA1	452.6	b
AIC_RA	477.9	c
Hypospadias		
AIC DA OBS	897.6	a
AIC_DA1	924.5	b
AIC DA2	928.9	b
AIC_RA	943.8	c
DA2=OBS 3 of 11		
DA2 > RA 10 of 11		
DA1>RA 9 OF 11		
DA2=DA1 7 of 11		
DA2>DA1 3 OF 11		
DA1>DA2 0 OF 11		

models with different letters differ from one another

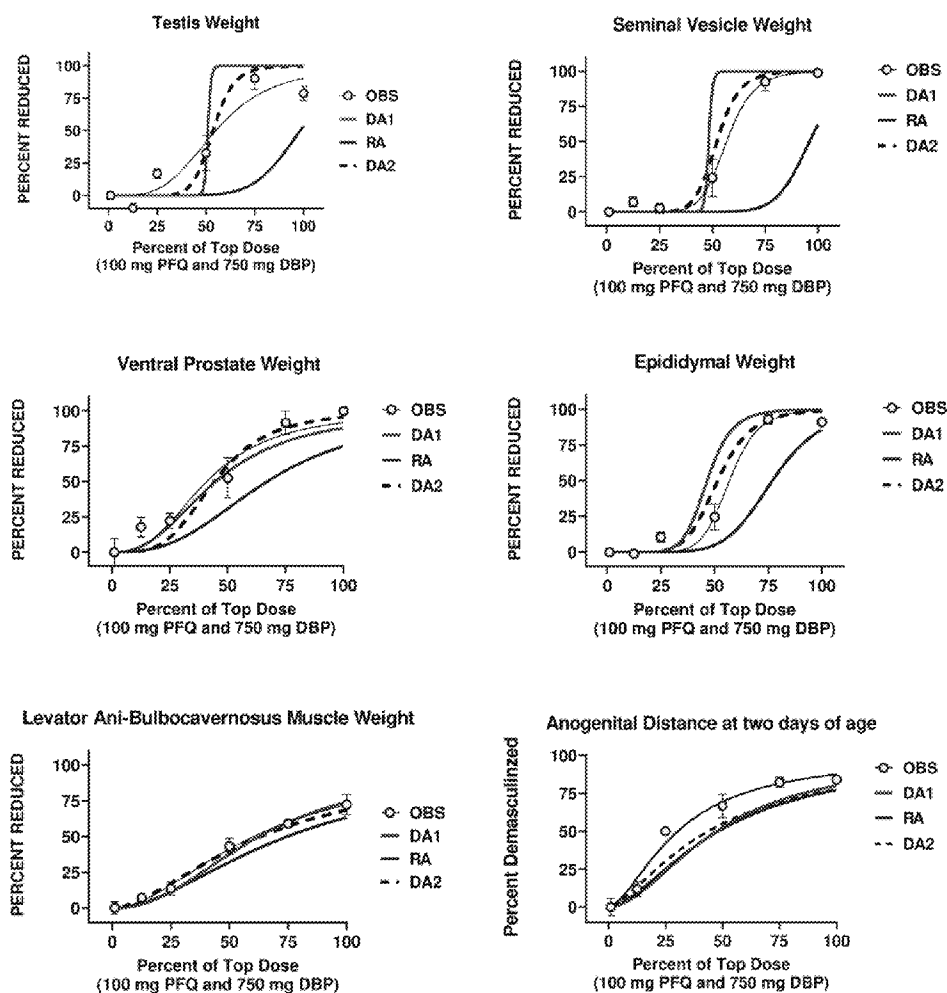


figure 1 a 4pl graphs

197x205mm (300 x 300 DPI)

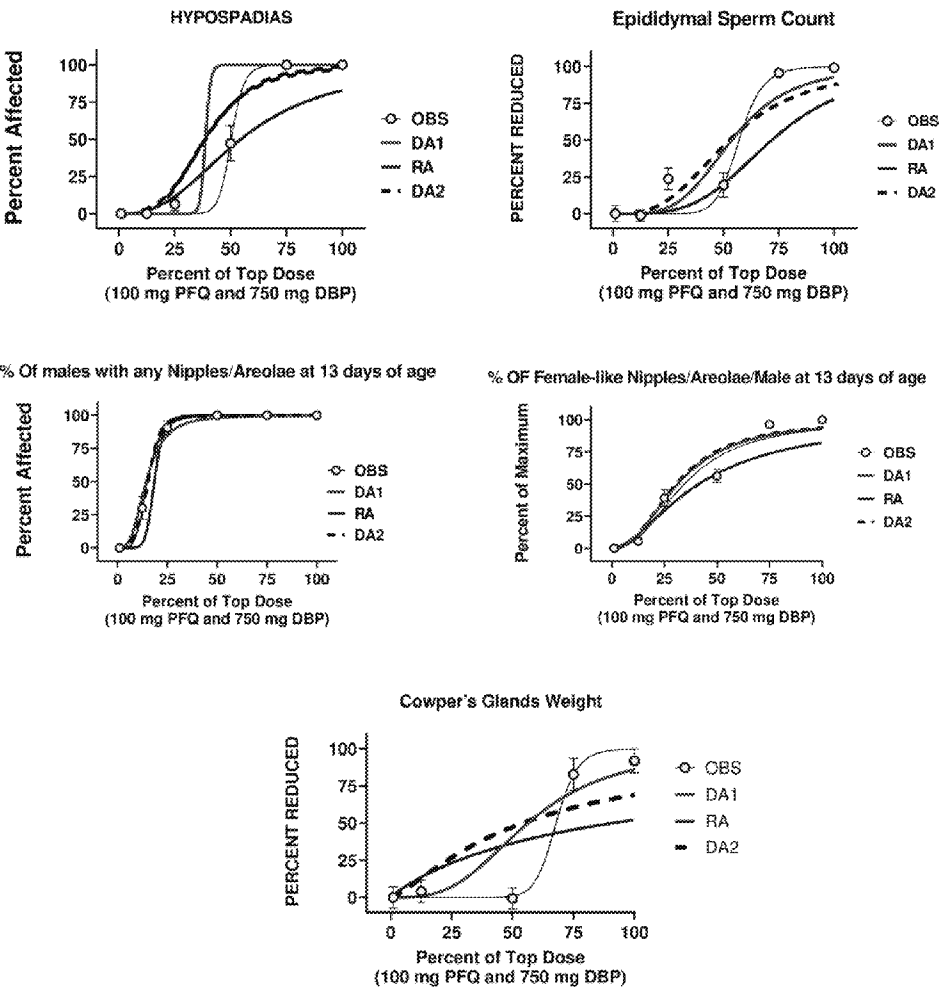


FIGURE 1B 4PL GRAPHS

199x207mm (300 x 300 DPI)

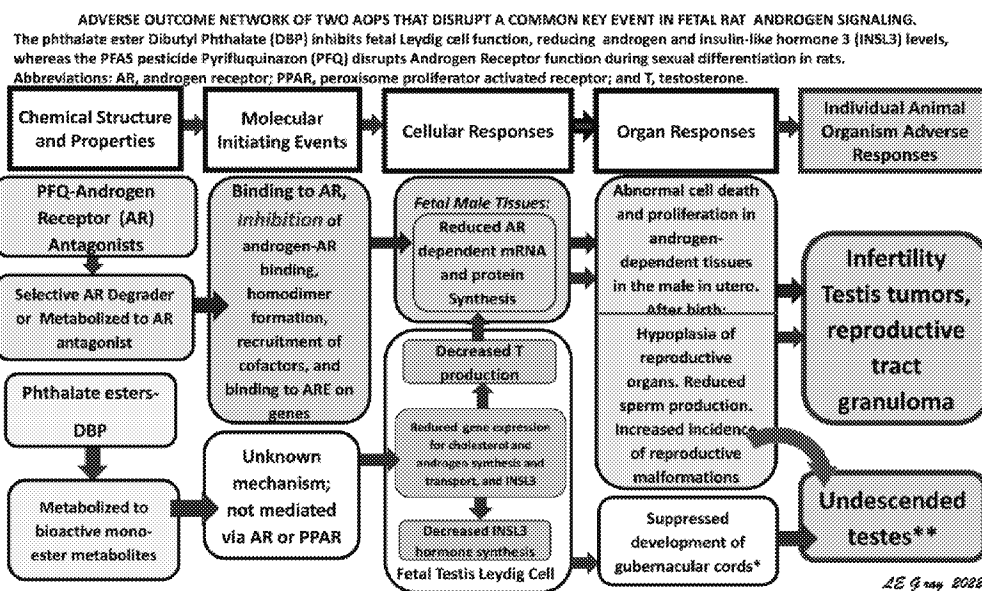


FIG 2 AOPn

228x137mm (300 x 300 DPI)

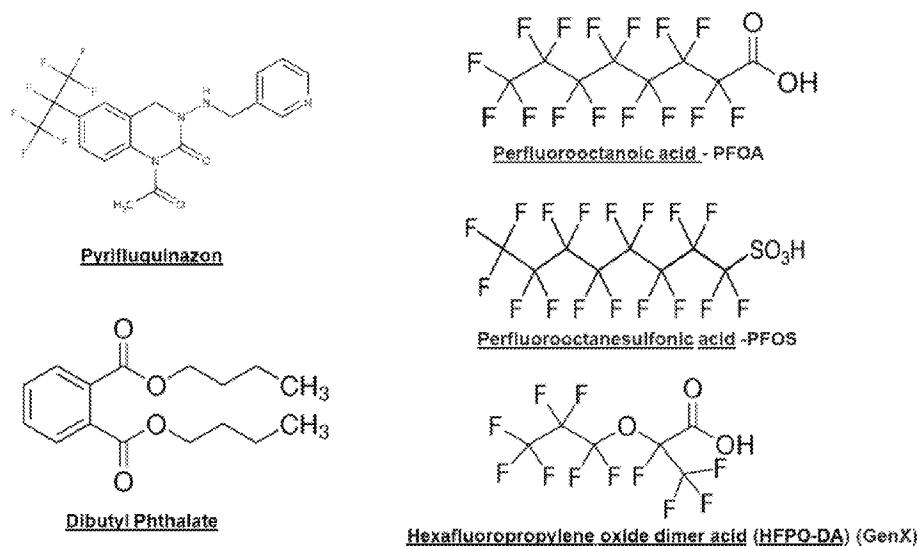


Fig 3 chemical structures

338x190mm (96 x 96 DPI)